# EFFECTS OF FLUORIDE ON C-REACTIVE PROTEIN, ADENOSINE DEAMINASE, AND CERULOPLASMIN IN RABBIT SERA

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SUMMARY: Twenty healthy 6-month-old male and female New Zealand rabbits with a mean body weight of 3.5±0.5 kg were fed a standard commercial rabbit diet and provided with water *ad libitum* containing 40 mg F/L for 70 days. Blood samples were obtained from each rabbit, and sera adenosine deaminase (ADA) activity and C-reactive protein (CRP) and ceruloplasmin (CP) levels were determined with the following results at the start, at day 35, and at day 70, respectively: ADA activity 9.55±0.66, 14.78±1.11, and 19.56±1.85 U/L; CRP level 18.1±2.6, 85.5±20.3, and 123±23.26 ng/mL; CP level 22.19±2.66, 19.49±1.18, and 14.75±0.96 mg/dL. By days 35 and 70 significant increases in ADA activity and CRP levels had occurred, along with a significant decrease in the CP level. These results demonstrated that fluoride intoxication caused significant alterations in ADA, CRP, and CP in rabbit sera.

Keywords: Adenosine deaminase, C-reactive protein; Ceruloplasmin; Fluoride in rabbits.

#### INTRODUCTION

Fluorosis is an important public health problem throughout many parts of the world. After chronic administration of fluoride to animals, various changes occur in the blood, brain, liver, muscle, heart, kidney, and spinal cord. These changes include abnormal behaviour patterns, altered neuronal and cerebrovascular integrity, and metabolic lesions. Generation of lipid peroxidation, free radicals, and altered antioxidant defence systems are considered to play an important role in the toxic effects of fluoride.<sup>1-7</sup> Exposure to fluorides can induce inflammatory reactions and cell cycle arrest<sup>8</sup> and an increase in the levels of markers of inflammatory reactions.<sup>9</sup> C-reactive protein (CRP) is synthesized in appreciable amounts following tissue injury, and is used as a marker of an inflammatory reaction.<sup>10</sup> Ceruloplasmin (CP) is produced by hepatocytes, and its synthesis is accelerated in response to inflammation.<sup>11</sup> Adenosine deaminase (ADA) also plays an important role in acute and protracted inflammatory responses.<sup>12</sup>

The aim of the present study was to investigate alterations in sera ADA activity and CRP and CP levels caused by elevated levels of fluoride ingestion from drinking water in rabbits.

#### MATERIALS AND METHODS

Animals and experimental procedure: Twenty healthy 6-month-old male and female New Zealand rabbits with a mean body weight of  $3.5\pm0.5$  kg were used. The rabbits were obtained from the Ondokuz Mayis University Animal Laboratory (Samsun, Turkey), and the study was approved by the University Ethics Committee.

After their clinical health was verified, the animals were housed in a well-ventilated, temperature-controlled  $(23\pm2^{\circ}C)$  hygienic room at 60% relative humidity under a 12-hr light/dark cycle. Throughout the study, the rabbits were

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allowed free access to standard rabbit chow (Samsun, Turkey) and drinking water containing 40 mg F/L. On day zero 7 mL of blood was collected from each rabbit via puncture of *v. auricularis magna* to determine the ADA activity and CRP and CP levels in the sera. Subsequently, on days 35 and 70, blood samples were again collected from each animal. All blood samples were placed in tubes and centrifuged (1550 g, 10 min, +4°C). The separated serum samples were then stored at  $-80^{\circ}$ C until analysed.

*Biochemical analyses:* Serum ADA activities were measured using the Giusti method.<sup>13</sup> CRP was analysed with diagnostic commercial Elisa kits (Life Diagnostics, Inc. 2210-5). Serum CP analysis was conducted by a spectrophotometric method, which included use of *p*-phenyldiamine dichloride (PPD).<sup>14</sup>

*Statistical analysis:* The significance of differences between pre- and post-testing results was determined using the paired *t*-test.

## RESULTS

Results of the analyses are given in the figure.

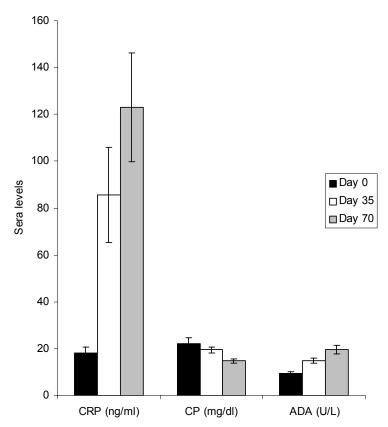


Figure. Rabbit sera CRP (ng/mL) activity, CP (mg/dL) levels and ADA (U/L) activity on days 0, 35, and 70(n=20).

As indicated by the data, a significant increase was observed in serum ADA activity (p<0.001) on days 35 and 70 and in the serum CRP level on day 35 (p<0.01) and 70 (p<0.001). For the CP level, however, a significant decrease was observed only on day 70 (p<0.01) compared to day 0.

## DISCUSSION

The role of fluoride as a possible activator in the monocyte differentiation process also seems to be confirmed by the results of *in vivo* studies.<sup>15</sup> In addition, fluoride acts as an activator of alveolar macrophages enhancing the production of chemokines and pro-inflammatory cytokines (pro-inflammatory activity).<sup>16</sup> Furthermore, epithelial lung cell exposure to fluoride had been shown to release increased amounts of inflammatory cytokines, and fluoride has been reported to increase the production of cytokine.<sup>17</sup>

C-reactive protein (CRP) is a sensitive marker of systemic low-grade inflammation and is currently recommended as the principal inflammatory marker in research and clinical practice.<sup>18</sup> In our work, significantly higher CRP levels were observed in rabbit sera on days 35 (p<0.01) and 70 (p<0.001, compared to the level on day 0. Although few studies on CRP levels in fluorosis are on record, our findings are in accord with previous results reported by Susheela and Jethanandani.<sup>9</sup>

Adenosine deaminase (ADA) acts in differentiating lymphoid cells and is secreted in biological fluids during the cellular immune response against intracellular pathogens, but it can also be increased in other pathological processes.<sup>19-23</sup> Macrophages have been suggested as the cellular source of extracellular ADA activity. ADA is important in acute and protracted inflammatory responses. Recent work in our laboratory has demonstrated elevated ADA activity during inflammatory responses in macrophage-rich tissues, such as liver and spleen.<sup>12,24,25</sup> Similarly, the present study revealed significantly elevated sera ADA activity (p<0.001).

Ceruloplasmin (CP) is produced by hepatocytes, and its synthesis is accelerated in response to inflammation.<sup>11</sup> In our work, significant decreases (p<0.01) in CP levels were observed in rabbit sera on day 70, compared to that on day 0. Although not many reports were encountered about CP values in fluoride intoxication, results of Sharma<sup>26</sup> are concordant with those of the present study. Previous studies have also indicated that copper levels in serum decreased after chronic fluorosis.<sup>27-29</sup> Since CP is a copper-containing serum protein, this decrease in CP level may be related to the reduction in the levels of serum copper.

In conclusion, statistically significant increases in CRP levels and in ADA activities, along with the decrease in CP levels by day 35 and 70 observed in our study suggest that the ingestion of fluoride by rabbits via drinking water at 40 mg F/L caused acute phase response for both study periods. The data also indicate that acute phase protein CRP and CP levels and ADA activities, along with other parameters, may be determinative criteria for diagnosing fluoride poisoning.

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### REFERENCES

- 1 Rzeuski R, Chlubek D, Machoy Z. Interactions between fluoride and biological free radical reactions. Fluoride 1998;31:43-5.
- 2 Sharma A, Chinoy NJ. Role of free radicals in fluoride-induced toxicity in liver and kidney of mice and its reversal [abstract]. Fluoride 1998;31:S26.
- 3 Kaul RD, Susheela AK. Evidence of muscle fiber degeneration in rabbits treated with sodium fluoride. Fluoride 1974;7:177-81.
- 4 Shashi A, Singh JP, Thapar SP. Protein degradation in skeletal muscle of rabbit during experimental fluorosis. Fluoride 1992;25:155-8.
- 5 Shashi A, Singh JP, Thapar SP. Effect of long-term administration of fluoride on levels of protein, free amino acids and RNA in rabbit brain. Fluoride 1994;27:155-9.
- 6 Singh M. Biochemical and cytochemical alterations in liver and kidney following experimental fluorosis. Fluoride 1984;17:81-93.
- 7 Guan ZZ, Yang PS, Yu ND, Zhuang ZJ. An experimental study of blood biochemical diagnostic indices for chronic fluorosis. Fluoride 1989;22:112-8.
- 8 Thrane EV, Refsnes M, Thoresen GH, Låg M, Schwarze PE. Fluoride-induced apoptosis in epithelial lung cells involves activation of MAP kinases p38 and possibly JNK. Toxicol Sci 2001;61:83-91.
- 9 Susheela AK, Jethanandani P. Serum haptoglobin and C-reactive protein in human skeletal fluorosis. Clin Biochem 1994;27:463-8.
- 10 Steele DM, Whitehead AS. The acute phase response. In: Sim E, ed. Humoral factors. New York: Oxford University Pres;1993.
- 11 Voelkel EF, Levine L, Alper CA, Tashjian AH Jr. Acute phase reactants ceruloplasmin and haptoglobin and their relationship to plasma prostaglandins in rabbits bearing the VS2 carcinoma. J Exp Med 1978;148:1078-88.
- 12 Adanin S, Yalovetskiy IV, Nardulli BA, Sam ADII, Jonjev ZS, Law WR. Inhibiting adenosine deaminase modulates the systemic inflammatory response syndrome in endotoxemia and sepsis. Am J Physiol 2002;282:1324-32.
- 13 Giuisti G. Adenosine deaminase. In: HU Bergmeyer, ed. Methods of enzymatic analysis. New York: Academic Press;1974.
- 14 Colombo JP, Richterich R. Zur bestimmung des caeruloplasmin in plasma [On the determination of ceruloplasmin in plasma]. Schweiz Med Wochenschr 1964;94:715-20. [in German].
- 15 Hirano S, Ando M. Apoptotic cell death following exposure to fluoride in rat alveolar macrophages. Arch Toxicol 1996;70: 249-51.
- 16 Stachowska E, Baśkiewicz-Masiuk M, Machaliński B, Rybicka M, Gutowska I, Bober J, et al. Sodium fluoride enhancement of monocyte differentiation via nuclear factor KB Mechanism. Fluoride 2005;38:297-306.
- 17 Refsnes M, Becher R, Låg M, Skuland T, Schwarze PE. Fluoride-induced interleukin-6 and interleukin-8 synthesis in human epithelial lung cells. Hum Exp Toxicol 1999;18:645-52.
- 18 Pearson TA, Mensah GA, Alexander RW, Anderson JL, Cannon RO, Criqui M, et al. Markers of inflammation and cardiovascular disease: application to clinical and public health practice: A statement for healthcare professionals from the Centers for Disease Control and Prevention and the American Heart Association. Circulation 2003;107:499-511.
- 19 Giusti G, Galanti B. Colorimetric method. In: HU Bergmeyer, Methods of Enzymatic Analysis, 3rd ed., Weinheim: Verlag Chemie; 1984. p. 315-23.
- 20 Edwards YH, Hopkinson DA, Harris H. Adenosine deaminase isozymes in human tissues. Ann Hum Genet 1971;35:207-19.
- 21 Bothamley GH. Tuberculous pleurisy and adenosine deaminase. Thorax 1995;40:593-4.
- 22 L'Herminez RH. Urgent need for a new approach tool the diagnosis of tuberculosis in developing countries in the decade of AIDS. Trop Geograp Med 1993;45:145-9.

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- 23 Chalhoub M, Cruz AA, Marcílio C, Netto MB. Valor da determinação da atividade de adenosina desaminase (ADA) no diagnóstico diferencial dos derrames pleurais [Value of determining the activity of adenosine deaminase (ADA) in the differential diagnosis of pleural effusions]. Rev Ass Med Brasil 1996;I42:139-46. [in Portuguese].
- 24 Cohen ES, Law WR, Easington CR, Kuruz KQ, Nardulli BA, Balk RA, et al. Adenosine deaminase inhibition attenuates microvascular dysfunction and improves survival in sepsis. Am J Resp Crit Care Med 2002;166:16-20.
- 25 Law WR, Conlon BA, Valli VE. Therapeutic potential for transient inhibition of adenosine deaminase in systemic inflammatory response syndrome. Crit Care Med 2003;31:1475-81.
- 26 Sharma YD. Serum sialic acid and ceruloplasmin levels in experimental fluorosis. Toxicol Lett 1983;15:1-5.
- 27 Kanwarand KC, Singh M. Zinc, copper and manganese levels in various tissues following fluoride administration. Cell Mol Life Sci 1981;37:1328-9.
- 28 Singh M, Kanwar KC. Copper and iron in tissue following experimental fluorosis, Fluoride 1981;14:107-12.
- 29 Bouaziz H, Croute F, Boudawara T, Soleilhavoup JP, Zeghal N. Oxidative stress induced by fluoride in adult mice and their suckling pups. Exp Toxicol Pathol 2007;58:349-59.