

## ALLEVIATORY EFFECTS OF HYDROALCOHOLIC EXTRACT OF CAULIFLOWER (*BRASSICA OLERACEA* VAR. *BOTRYTIS*) ON THYROID FUNCTION IN FLUORIDE INTOXICATED RATS

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**ABSTRACT:** Fluorosis is a progressive degenerative disorder. Excess intake of fluoride (F) affects the functioning of the thyroid gland. The present study was conducted to evaluate the alleviatory effect of a hydroalcoholic extract of leaves of the cauliflower plant on thyroid function in sodium fluoride (NaF) intoxicated rats. Thirty-six male Wistar albino rats were divided into six groups of six animals in each: Group I served as the normal control; Group II served as the toxic control; Groups III, IV, and V served as treatment groups and received extract at three doses 100 mg, 200 mg, and 400 mg per kg body weight respectively; and Group VI served as the plant control and received a hydroalcoholic extract of leaves of cauliflower plant in a dose of 400 mg per kg body weight. All groups except I and VI received NaF (100 ppm) through drinking water for 30 days. After completion of the study, the animals were fasted overnight, blood was collected by retro-orbital puncture, and the serum levels of thyroxine (T<sub>4</sub>) and triiodothyronine (T<sub>3</sub>) were measured by radioimmunoassay. Compared to the toxic control group, the administration of the 100 and 200 mg doses of the hydroalcoholic extract of cauliflower leaves resulted significant alleviatory reductions in the levels of total T<sub>4</sub> and the administration of the 200 and 400 mg doses of the extract resulted in significant alleviatory reductions in the levels of total T<sub>3</sub>.

Keywords: Cauliflower plant (*Brassica oleracea* var. *botrytis*); Sodium fluoride; Thyroid hormones.

### INTRODUCTION

The thyroid hormones, triiodothyronine (T<sub>3</sub>) and thyroxine (T<sub>4</sub>) are tyrosine-based hormones produced by the follicular cells of the thyroid gland and are regulated by thyroid-stimulating hormone (TSH) made by the thyrotropes of the anterior pituitary gland (adenohypophysis). Both the hormones are essential for the proper development and differentiation of all the cells of the human body, and the regulation of protein, fat, and carbohydrate metabolism. Thyroid hormones increase heart rate, cardiac contractility, and cardiac output. They also promote vasodilation, which leads to enhanced blood flow to many organs. In addition, thyroid also produces calcitonin, which plays a vital role in calcium (Ca<sup>2+</sup>), and phosphorus metabolism.<sup>1</sup>

Fluorine is a highly reactive element which does not exist in a free form in nature but instead is commonly present as the fluoride ion (F<sup>-</sup>). The main sources of F intake include drinking water, soil, food, dental products, industrial dust, and smoke. It is not an essential trace element and is not required for the healthy development of teeth or bones. Since the late 1940s the use of fluorides for the

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prevention of dental caries—especially the adjustment of the fluoride content of drinking water—has been a subject of considerable controversy.<sup>2</sup> In higher doses fluoride may cause dental, skeletal, and nonskeletal fluorosis in both human beings<sup>3-6</sup> and domestic animals.<sup>7-10</sup> Experimentally, it has also been observed and reported that chronic exposure to a relatively low level of F causes adverse changes in diverse soft tissues and organs including thyroid dysfunction.<sup>11</sup>

Globally, many plant based nutritional supplements and foods are recognized for having therapeutic effects on some chronic and degenerative diseases with fewer side effects than some synthetic medications. Many nutritionally rich plants have the potential for ameliorating F-induced toxicity.<sup>12-20</sup>

Cauliflower (*Brassica oleracea* var. *botrytis*) is one of the economical, nutritionally rich plants and contains flavonoids and polyphenolic compounds. Its claimed properties include anticancer, antiobesity, antioxidant, anti-inflammatory agent, and hepatoprotective effects.<sup>21</sup> No previous studies have been done of the possible protective effects of cauliflower leaves on NaF-induced toxicity. In the present study, we investigated the effect of a hydroalcoholic extract of cauliflower leaves on F-induced thyroid gland dysfunction.

#### MATERIALS AND METHODS

*Collection and authentication of plant material:* Fresh leaves of cauliflower were collected from a local market in Kadapa, Kadapa district, Andhra Pradesh, India, during the month of June, 2014. The plant material was authenticated by the Chief Scientist, Raw Material Herbarium and Museum, Delhi, and a certified specimen was stored in the Department of Pharmacognosy, Ch. Malla Reddy College of Pharmacy, Hyderabad, India.

*Preparation of extract:* Initially, the freshly collected leaves were cleaned and shade dried. After drying, the leaves were milled into a coarse powder by a mechanical grinder (sieve no: 10/44). The coarse powder was extracted with hydro-alcohol (30:70) by using a maceration process. The solvent was removed under reduced pressure to get a semisolid residue (yield 31.4% w/w). The extract was stored in an airtight container at 4°C for further studies.

*Animals:* Male Wistar rats weighing 200–250 g were used for the study. The animals were obtained from Sai Tirumala Enterprises Pvt Ltd, Hyderabad, India. The animals were randomly allocated to the treatment groups in polypropylene cages with paddy husk as bedding. The animals were housed at a temperature of 24±2°C. A 12:12 hr light:dark cycle was followed. They were fed with standard commercial pellets (containing total carbohydrates 54%, crude protein 19.96%, crude fat 3.73%, crude fiber 3.50%, calcium 1.44%, and phosphorus 0.81%) rat chow, and water *ad libitum* during the experiment. All the experimental procedures and protocols used in this study were reviewed by the Institutional Animal Ethics Committee (IAEC) and were in accordance with the guidelines of the IAEC (IAEC no: CPCSSSEA/1657/IAEC/CMRCP/PhD-14/35).

*Experimental design:* The dose of NaF was selected based on the previous studies,<sup>14</sup> and after ten days of adaptation period, thirty-six animals were divided into six groups of six animals each. Group I served as a normal control and

received drinking water. Group II served as a toxic control and received NaF (100 ppm) through drinking water for 30 days. Groups III, IV, and V served as the treatment groups and received the cauliflower leaves extract (100 mg/kg, 200 mg/kg, and 400 mg/kg, respectively) for 30 days and NaF administered through drinking water. Group VI served as a plant control and received cauliflower leaves extract alone (400 mg/kg) once daily for 30 days. After the treatment schedule, animals were fasted overnight and blood, collected by retro-orbital puncture, was allowed to clot for approximately 1 hr at room temperature and then centrifuged at 2,500 rpm for 15 min to obtain the serum.

*Estimation of thyroxine and triiodothyronine:* The serum thyroxine and triiodothyronine levels were estimated by radioimmunoassay using a commercially available Cis-Bio RIA Kit.

*The statistical analysis* was done using GraphPad InStat™, version 3.10.<sup>22</sup>

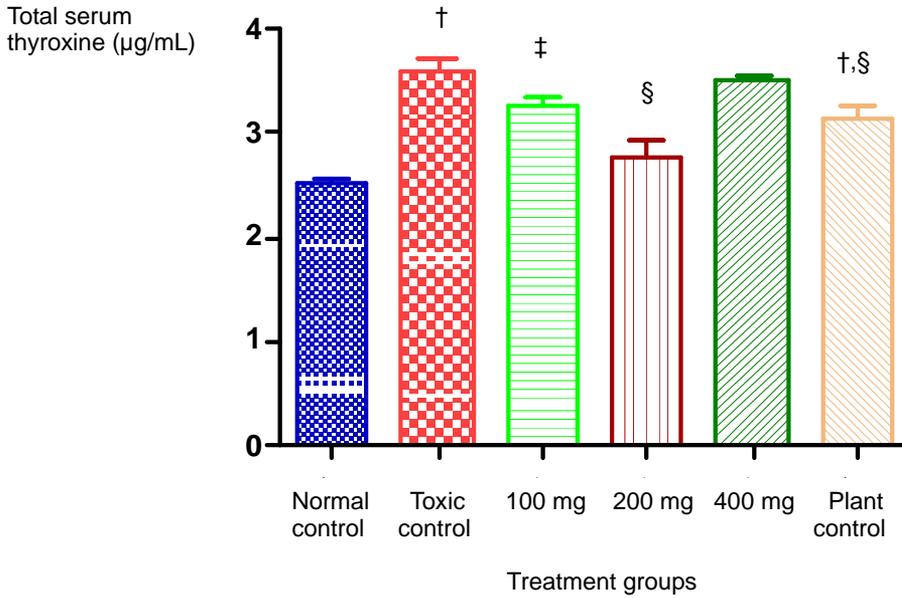
### RESULTS

The NaF-intoxicated rats showed increased serum levels of total T<sub>4</sub> and total T<sub>3</sub> compared to the normal control group (Table 1). Compared to the toxic control group, the administration of the 100 and 200 mg doses of the hydroalcoholic extract of cauliflower leaves resulted significant reductions in the levels of total T<sub>4</sub> and the administration of the 200 and 400 mg doses of the extract resulted in significant reductions in the levels of total T<sub>3</sub>. (Figures 1 and 2). Compared to the normal control group, the administration of 400 mg of the hydroalcoholic extract of cauliflower leaves alone resulted in a significant increase in the level of T<sub>4</sub> but no significant change in the T<sub>3</sub> level. The results clearly showed that the hydroalcoholic extract of cauliflower leaves stimulated thyroid function.

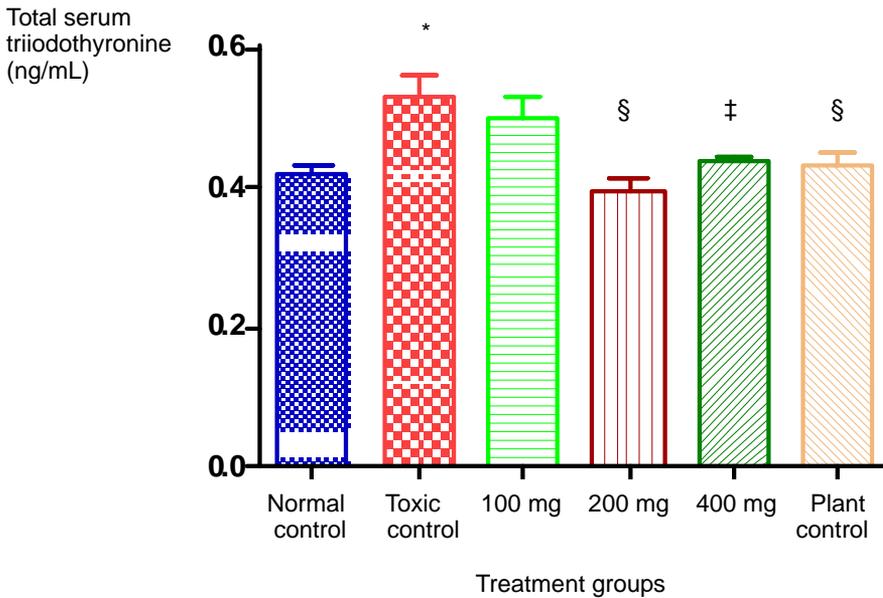
**Table 1.** Effects of hydroalcoholic extract of cauliflower leaves on total serum thyroxine and triiodothyronine in sodium fluoride intoxicated rats

Group	Total serum thyroxine±SD (µg/mL)	Total serum triiodothyronine±SE (ng/mL)
I Normal control	2.52±0.06	0.42±0.01
II Toxic control, 100 ppm NaF	3.60±0.12 <sup>†</sup>	0.53±0.03*
III 100 mg cauliflower leaves extract + 100 ppm NaF	3.27±0.08 <sup>‡</sup>	0.5±0.03
IV 200 mg cauliflower leaves extract + 100 ppm NaF	2.76±0.17 <sup>§</sup>	0.39±0.02 <sup>§</sup>
V 400 mg cauliflower leaves extract + 100 ppm NaF	3.49±0.04	0.44±0.01 <sup>‡</sup>
VI Plant control, 400 mg cauliflower leaves extract	3.15±0.11 <sup>†,§</sup>	0.43±0.01 <sup>§</sup>

Compared to the normal control group: \*p<0.001, <sup>†</sup>p<0.0001; compared to toxic control group: <sup>‡</sup>p<0.001, <sup>§</sup>p<0.0001



**Figure 1.** Effects of hydroalcoholic extract of cauliflower leaves on total serum thyroxine level in sodium fluoride intoxicated rats. Compared to the normal control group: \* $p < 0.001$ , † $p < 0.0001$ ; compared to toxic control group: ‡ $p < 0.001$ , § $p < 0.0001$ .



**Figure 2.** Effects of hydroalcoholic extract of cauliflower leaves on total serum triiodothyronine level in sodium fluoride intoxicated rats. Compared to the normal control group: \* $p < 0.001$ , † $p < 0.0001$ ; compared to toxic control group: ‡ $p < 0.001$ , § $p < 0.0001$ .

## DISCUSSION

Fluorine is a halogen chemically related to iodine but is chemically more active than iodine. The fluoride ion is a cumulative poison and can interfere with iodine metabolism in the thyroid gland by having an antagonistic effect. F accumulates in the thyroid gland and can directly injure the structure of thyroid follicle, provoke cytoplasm reduction and karyopyknosis of follicular epithelial cells, diminish the count of microvilli on the cristae of epithelial cells, and lead to bulging of the vacuoles in the follicular epithelial cells.<sup>23</sup> Earlier studies conducted in both animals and human beings clearly indicated that the thyroid gland is sensitive to the F.<sup>24-29</sup>

A possible mechanism for the increased serum level of total T<sub>4</sub> and total T<sub>3</sub> might be an activation by F of the pituitary gland resulting in the release of more T<sub>4</sub> from follicular cells. In turn, the T<sub>4</sub> is then deiodinated and changed into active T<sub>3</sub> by deiodinase in order to supply T<sub>3</sub> in the blood circulation.<sup>30</sup>

Many effects primarily attributed to fluoride are caused by the synergistic action of fluoride plus aluminum.<sup>31</sup> Soluble aluminofluoride complexes, fluoroaluminate (AlF<sub>4</sub>), are formed in water solutions containing fluoride and traces of aluminum.<sup>32</sup> These complexes are able to simulate phosphate groups in many biochemical reactions.<sup>32</sup> AlF<sub>x</sub> are used in many laboratory investigations of guanine nucleotide proteins (G proteins).<sup>32</sup> They affect various enzyme activities and cell signalling cascades.<sup>32</sup> Fluoride in a daily dose of 5–10 mg/day has been used to treat hyperthyroidism.<sup>33</sup> It has been hypothesized that fluoride mimics the thyroid stimulating hormone (TSH) and although this would stimulate thyroid function rather than lower it the overproduction of cyclic adenosine monophosphate (cAMP) results in a desensitization of the TSH receptor leading ultimately to reduced activity of the thyroid gland.<sup>32,34</sup>

Susheela et al. found thyroid hormone derangements in 49 of 90 children (54.4%) living in fluoride endemic, non-iodine deficient areas with five patterns of disturbance: (i) high TSH with normal free T<sub>4</sub> (FT<sub>4</sub>) and free T<sub>3</sub> (FT<sub>3</sub>) (46.9%); (ii) normal TSH and FT<sub>4</sub> but low FT<sub>3</sub> (32.7%); (iii) high TSH and FT<sub>3</sub> with normal FT<sub>4</sub> (14.3%); (iv) high TSH with normal FT<sub>3</sub> but low FT<sub>4</sub> (4.1%); and (v) high TSH with normal FT<sub>4</sub> but low FT<sub>3</sub> (2.0%).<sup>28</sup> They noted that a negative feedback mechanism is involved in the production of thyroid hormones so that when the pituitary gland senses a drop in circulating FT<sub>3</sub> levels it releases more TSH to stimulate the production of the “pro-hormone” T<sub>4</sub> which then undergoes peripheral deiodination to produce the active hormone T<sub>3</sub>.<sup>28</sup> Three iodothyronine deiodinase enzymes, D<sub>1</sub>, D<sub>2</sub>, and D<sub>3</sub>, catalyse the deiodination. D<sub>1</sub>, expressed in the thyroid gland, liver, and kidney, converts T<sub>4</sub> to T<sub>3</sub> in peripheral tissues, particularly in the liver, and is reflected in plasma T<sub>3</sub> levels. D<sub>2</sub>, expressed in brain, pituitary gland, and skeletal muscle, converts T<sub>4</sub> to T<sub>3</sub> in local tissues, and through its presence in skeletal muscle makes some contribution to the plasma T<sub>3</sub>.<sup>28</sup> D<sub>3</sub>, expressed in brain, placenta, and fetal tissues, is an inactivating enzyme producing inactive metabolites by converting T<sub>4</sub> to reverse T<sub>3</sub> (rT<sub>3</sub>) and T<sub>3</sub> to 3,3'-T<sub>2</sub>.<sup>28</sup>

The release of TSH is controlled by positive input from the hypothalamic hormone thyrotropin-releasing hormone (TRH) and by negative input from  $T_4$  and  $T_3$ , with the  $T_3$  rather the  $T_4$  being responsible for the feedback response.<sup>35</sup> TSH binds to G-protein-coupled receptors in the surface membranes of thyroid follicular cells which leads to increases in both cAMP and diacylglycerol/inositol triphosphate second messenger pathways.<sup>35</sup> Some  $T_3$ , the active form of thyroid hormone, is secreted directly by the thyroid along with  $T_4$ , but most  $T_3$  is produced from  $T_4$  by one of two deiodinases ( $D_1$  and  $D_2$ ) in the peripheral tissue.  $T_3$  enters the nucleus of the target cells and binds to specific receptors, which activate specific genes.<sup>35</sup> Fluoride does not compete with iodide for transport into the thyroid.<sup>35</sup> The administration of fluoride in drinking water to rats results in variable effects on the  $T_4$  and  $T_3$ .<sup>35</sup> Hara found elevated  $T_3$  and  $T_4$  at the lowest dose of approximately 0.1 mg/kg/day.<sup>35,36</sup> With intermediate doses (3–4 mg/day) the  $T_3$  was decreased and the  $T_4$  was normal.<sup>35,36</sup> At the highest doses (10–20 mg/kg/day), TSH and growth hormone were decreased indicating possible effects on pituitary function.<sup>35,36</sup>

### CONCLUSION

In conclusion, this study has demonstrated that a hydroalcoholic extract of cauliflower leaves showed a significant alleviatory effect at doses of 100–400 mg/kg against NaF-induced thyroid gland dysfunction. However, further studies are required to understand the exact pathway or mechanism for the thyroregulatory role of the extract.

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### REFERENCES

- 1 El-Mehi AE, Amin SA. Effect of lead acetate on the thyroid gland of adult male albino rats and the possible protective role of zinc supplementation: a biochemical, histological and morphometric study. *J American Sci* 2012;8(7):61-71.
- 2 World Health Organization. Fluorides and human health. Geneva: World Health Organization; 1970.
- 3 Choubisa SL, Choubisa DK, Joshi SC, Choubisa L. Fluorosis in some tribal villages of Dungarpur district of Rajasthan, India. *Fluoride* 1997;30(4):223-8.
- 4 Choubisa SL. Endemic fluorosis in southern Rajasthan (India). *Fluoride* 2001;34(1):61-70.
- 5 Choubisa SL, Choubisa L, Choubisa D. Osteo-dental fluorosis in relation to nutritional status, living habits and occupation in rural areas of Rajasthan, India. *Fluoride* 2009;42(3):210-15.
- 6 Choubisa SL. Fluoride in drinking water and its toxocosis in tribals, Rajasthan, India. *Proc Natl Acad Sci India Sect B Biol Sci* 2012;82(2):325-30.
- 7 Choubisa SL. Osteo-dental fluorosis in horses and donkeys of Rajasthan, India. *Fluoride* 2010;43(1):5-10.
- 8 Choubisa SL. Fluorosis in dromedary camels of Rajasthan, India. *Fluoride* 2010;43(3):194-9.
- 9 Choubisa SL, Modasiya V, Bahura CK, Sheikh Z. Toxicity of fluoride in cattle of the Indian Thar Desert, Rajasthan, India. *Fluoride* 2012;45(4):371-6.
- 10 Choubisa SL. Fluoride toxicosis in immature herbivorous domestic animals living in low fluoride water endemic areas of Rajasthan, India: an observational survey. *Fluoride* 2013;46(1):19-24.
- 11 Choubisa SL. Status of fluorosis in animals. *Proc Natl Acad Sci India Sect B Biol Sci* 2012;82(3):331-9.

- 12 Narasimhachary AVR, Vasant RA. Medicinal plants as antioxidant agents: understanding their mechanism of action and therapeutic efficacy. 1st ed. Kerala: Research Signpost; 2012. pp 267-87.
- 13 Nabavi SF, Eslami SH, Moghaddam AH, Nabavi SM. Protective effects of curcumin against fluoride-induced oxidative stress in the rat brain. *Neurophysiology* 2011;43(4):287-91.
- 14 Vasant RA, Narasimhacharya AVR. Alleviatory effects of *Emblica officinalis* G. as a food supplement in fluoride induced hyperlipemia and oxidative stress. *Int J Pharm Pharm Sci* 2012;4(1):404-8.
- 15 Bhatt RN, Vyas DD, Meda RB, Rao MV. Mitigating effects of triphala on fluoride blood toxicity in rat. *Indian J Appl Res* 2013;3(11):553-5.
- 16 Ranjan R, Swarup D, Patra RC, Chandra V. *Tamarindus indica* L. and *Moringa oleifera* M. extract administration ameliorates fluoride toxicity in rabbits. *Indian J Exp Biol* 2009;47: 900-05.
- 17 Srilatha K, Banji D, Banji OJF, Vinod KR, Saidulu A. Investigation on the anti-genotoxic effect of *Ocimum sanctum* in fluoride induced genotoxicity. *Int Res J Pharm* 2013;4(5):160-64.
- 18 Choubisa SL, Mishra GV, Sheikh Z, Bhardwaj B, Mali P, Jaroli VJ. Food, fluoride, and fluorosis in domestic ruminants in the Dungarpur district of Rajasthan, India. *Fluoride* 2011;44(2):70-6.
- 19 Choubisa SL. Why desert camels are least afflicted with osteo-dental fluorosis? *Curr Sci* 2013;105(12):1671-2.
- 20 Choubisa SL, Mishra GV. Fluoride toxicosis in bovines and flocks of desert environment. *Int Pharmacol Biol Sci* 2013;7(3):35-40.
- 21 Raghavendra M, Reddy KR, Yadav PR, Jayaveera KN, Sowmyalatha A, Khan AR. Evaluation of acute toxicity and hepatoprotective effect of *Brassica oleracea* var. *botrytis* in experimental animals. *J Pharm Chem* 2014;8(2):12-6.
- 22 Motulsky HJ. GraphPad Prism 6™. San Diego, CA, USA: GraphPad Software, Inc; 2013. URL: <http://www.graphpad.com/company/>
- 23 Preedy VR, Burrow GN, Watson RR. Comprehensive handbook of iodine: nutritional, biochemical, pathological and therapeutic aspects. 1st ed. London: Elsevier Academic Press; 2009. pp 646-7.
- 24 Ge YM, Ning HM, Wang SL, Wang JD. DNA damage in thyroid gland cells of rats exposed to long-term intake of high fluoride and low iodine. *Fluoride* 2005;38(4):209-14.
- 25 Bouaziz H, Soussia L, Guermazi F, Zeghal N. Fluoride induced thyroid proliferative changes and their reversal in female mice and their pups. *Fluoride* 2005;38(3):185-92.
- 26 Zhan XA, Li JX, Wang M, Xu ZR. Effects of fluoride on growth and thyroid function in young pigs. *Fluoride* 2006;39(2):95-100.
- 27 Cinar A, Selcuk M. Effects of chronic fluorosis on thyroxine, triiodothyronine, and protein-bound iodine in cows. *Fluoride* 2005;38(1):65-8.
- 28 Susheela AK, Bhatnagar M, Vig K, Mondal NK. Excess fluoride ingestion and thyroid hormone derangements in children living in Delhi, India. *Fluoride* 2005;38(2):98-108.
- 29 Yasmin S, Ranjan S, Hilaluddin, Souza DD. Effect of excess fluoride ingestion on human thyroid function in Gaya region, Bihar, India. *Toxico Environ Chem* 2013;95(7):1235-43.
- 30 Ge YM, Ning HM, Gu XL, Yin M, Yang XF, Qi YH, Wang JD. Thyroid function in rats on high fluoride, low iodine. Feeding effects of high fluoride and low iodine on thyroid function in offspring rats. *J Integr Agric* 2013;12(3):502-8.
- 31 Strunecka A, Blaylock RL, Patocka J. Aluminofluoride complexes: phosphate analogues and a hidden hazard for living organisms. *Current Inorganic Chemistry* 2012;2:8-18.
- 32 Strunecka A, Patocka J, Blaylock RL, Chinoy NJ. Fluoride interactions: from molecules to disease. *Current Signal Transduction Therapy* 2007;2(3):190-213.
- 33 Galletti P-M, Joyet G. Effect of fluorine on thyroidal iodine metabolism in hyperthyroidism. *J Clin Endocrinol Metab* 1958;18(10):1102-10.
- 34 Tezelman S, Shaver JK, Grossman RF, Liang W, Siperstein AE, Duh QY, Clark OH. Desensitization of adenylate cyclase in Chinese hamster ovary cells transfected with human thyroid-stimulating hormone receptor. *Endocrinology* 1994;134(3):1561-9.
- 35 Doull J, Boekelheide K, Farishian BG, Isaacson RL, Klotz JB, Kumar JV, Limeback H, Poole C, Puzas JE, Reed N-MR, Thiessen KM, Webster TF, Committee on Fluoride in Drinking Water, Board on Environmental Studies and Toxicology, Division on Earth and Life Studies, National Research Council of the National Academies. Fluoride in drinking water: a scientific review of EPA's standards. Washington, DC: The National Academies Press; 2006. pp.224-6.
- 36 Hara K. Studies on fluorosis, especially effects of fluoride on thyroid metabolism. *Koku Eisei Gakkai Zasshi* 1980;30(10):42-57. [in Japanese].